

## Erk signaling activity is enhanced during collective migration on corrugated substrates

Zoé Barbier<sup>1</sup>, Tsuyoshi Hirashima<sup>2</sup>, Marine Luciano<sup>1</sup>, Sylvain Gabriele<sup>1\*</sup>

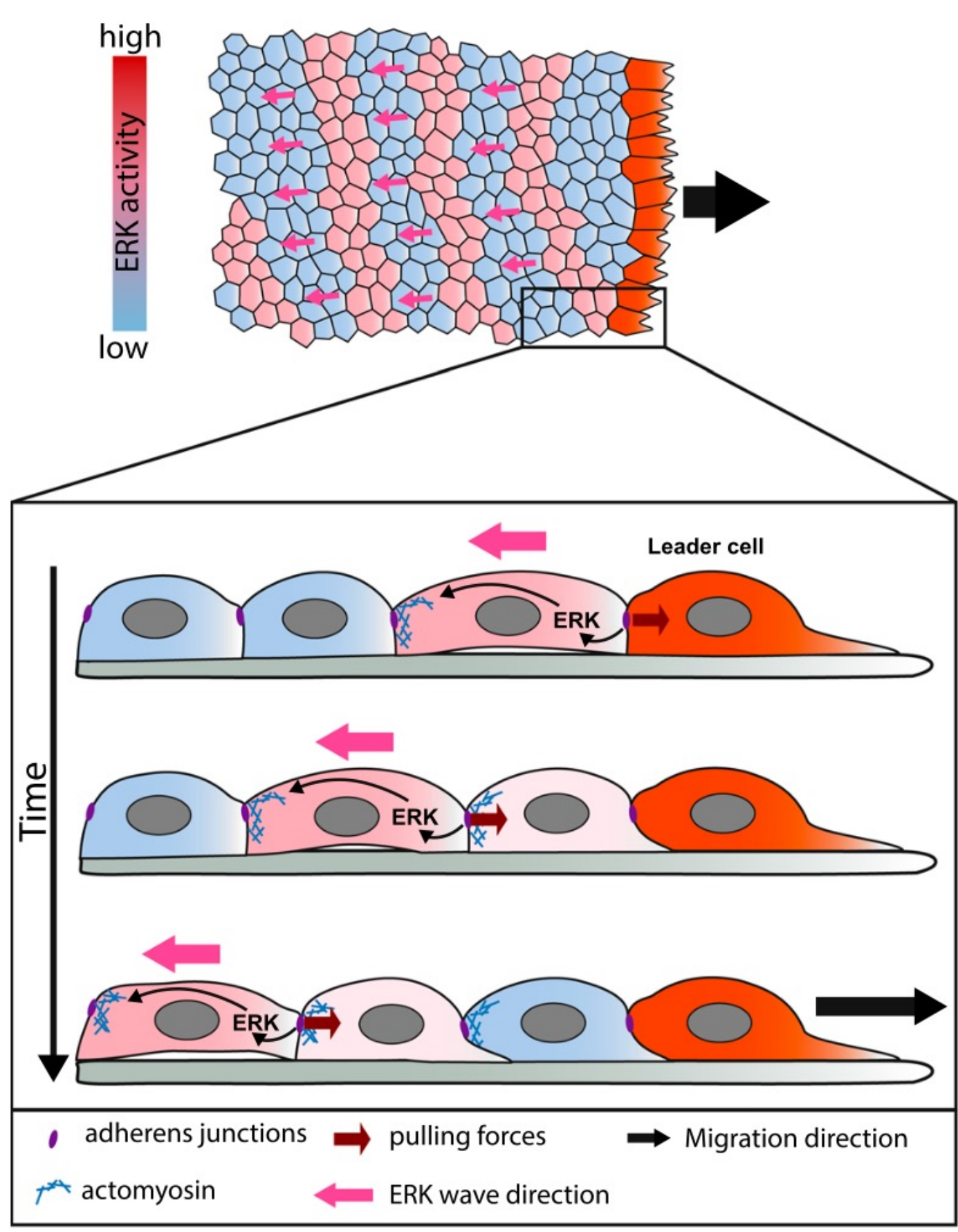
<sup>1</sup> Mechanobiology & Biomaterials Laboratory, Research Institute for Biosciences, CIRMAP, University of Mons, Place du Parc 20, B-7000 Mons, Belgium

<sup>2</sup>Mechanobiology Institute, National University of Singapore, 5A Engineering Drive 1, Level 9 T-Lab, Singapore 117411

\*Contact: [sylvain.gabriele@umons.ac.be](mailto:sylvain.gabriele@umons.ac.be)

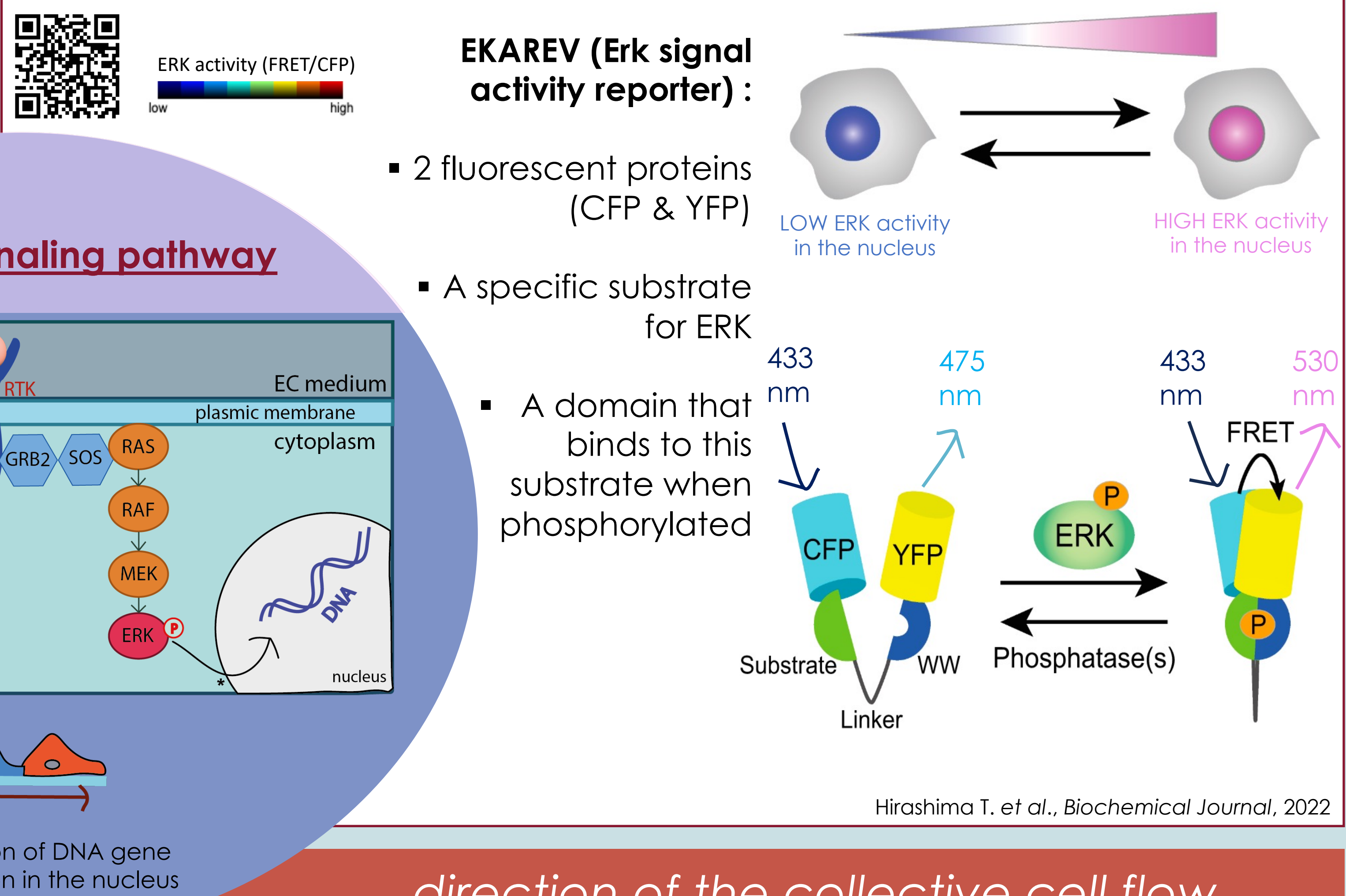
Collective migration plays a pivotal role in numerous epithelial tissues, under both physiological and pathological conditions. Recent evidence suggests a correlation between the activation waves of extracellular signal-regulated kinase (ERK) and cell deformation, which together orchestrate collective cell migration. Additionally, emerging evidence indicates that localized changes in matrix curvature can influence the migration of individual epithelial cells (known as curvotaxis). However, the precise mechanism by which local changes in curvature modulate ERK propagation and contribute to coordinating collective movement remains unclear. Here we utilize a photopolymerization technique to create well-defined corrugation patterns of varying wavelengths in soft hydrogels, thereby mimicking the multiscale curvature found in human tissues. To elucidate the role of ERK waves in directing cell migration, we employ Förster resonance energy transfer (FRET)-based biosensor in MDCK cells, conducting time-lapse experiments lasting 24 hours on culture substrates featuring both flat and corrugated regions. Our findings reveal distinct flow directions within the same epithelial tissue, transitioning from flat to corrugated hydrogels. Furthermore, we demonstrate that ERK activity is significantly heightened in corrugated regions, suggesting a mechanism that facilitates collective cell migration on curved substrates.

*ERK activity couples to polarization to direct collective cell migration*



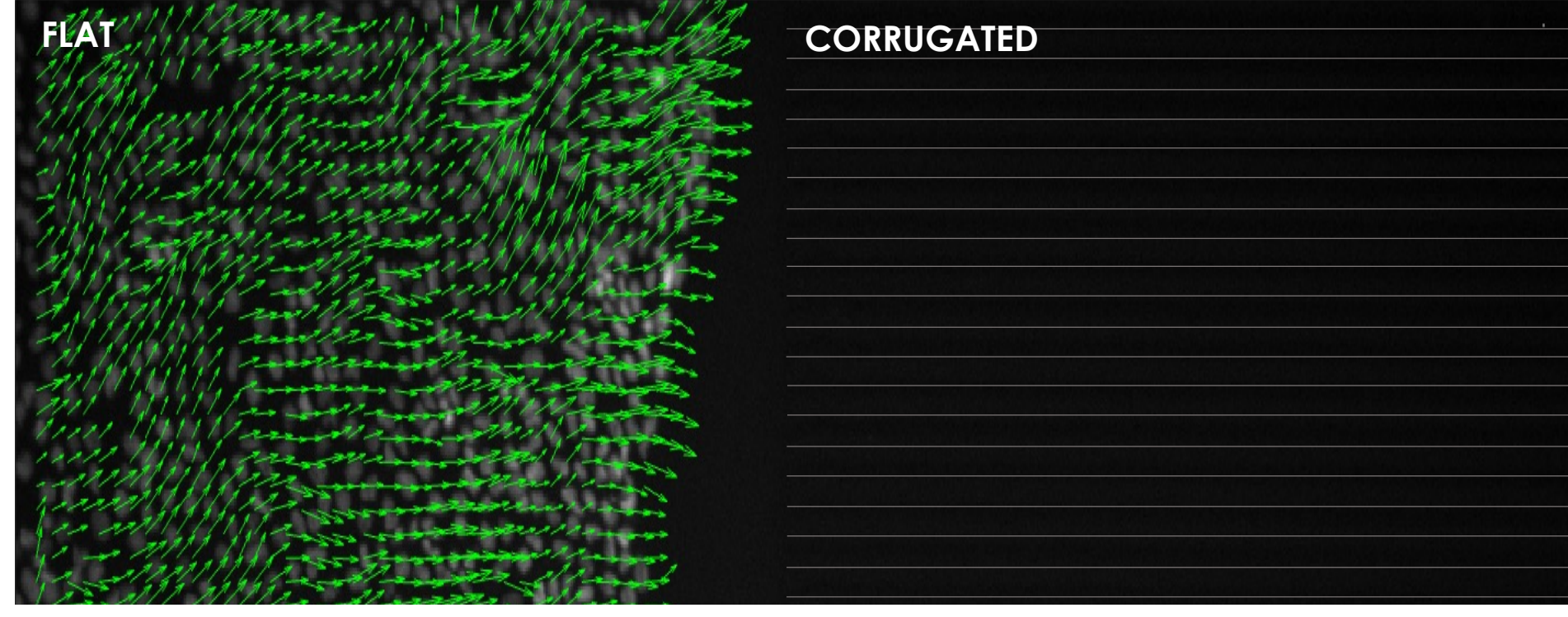
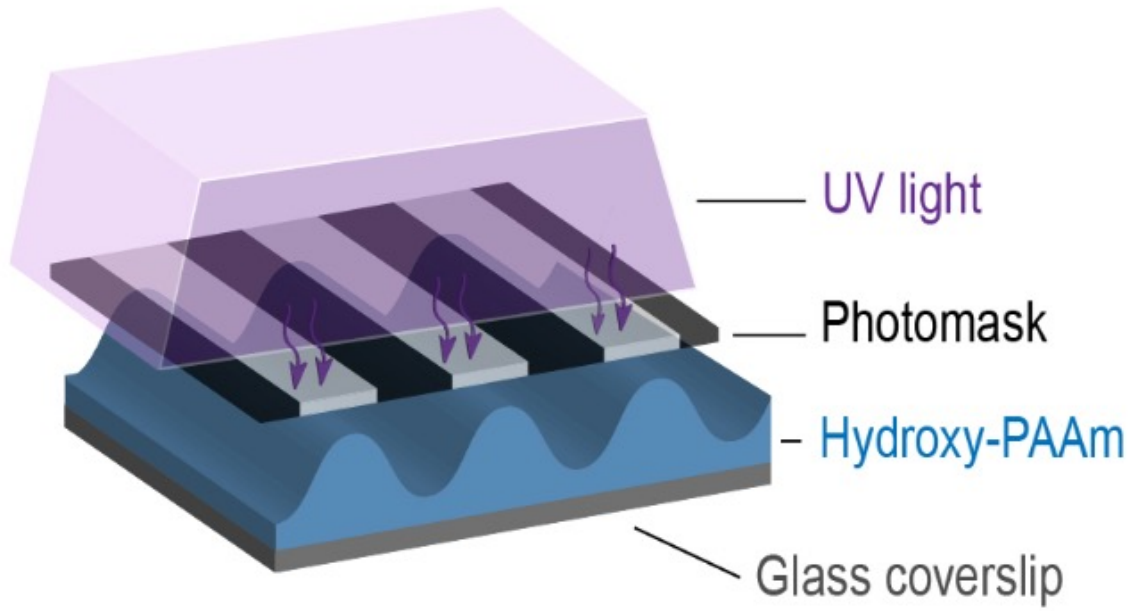
- Cell stretching promotes ERK activity via mechanical forces through adherens junctions
- ERK waves propagate towards the back side in a opposite direction of migration
- High-persistent ERK activity is present at the leader edge

## ERK activity is quantified by FRET biosensor

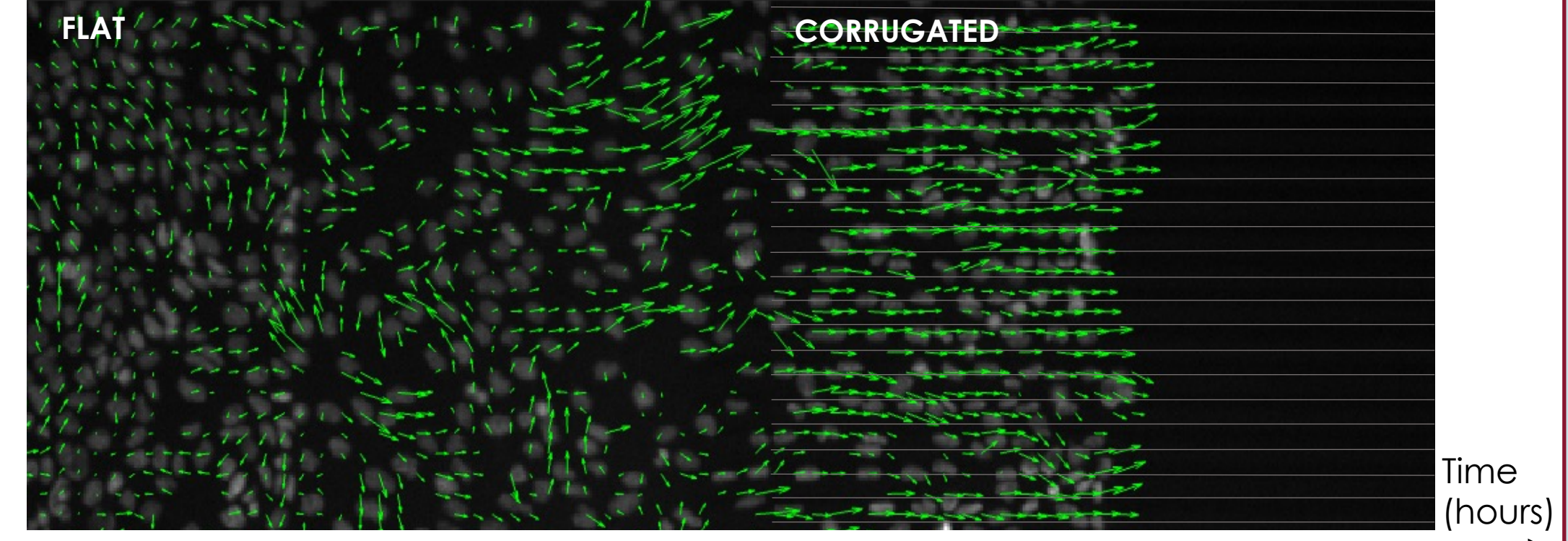
Hirashima T. et al., *Biochemical Journal*, 2022

## Substrate corrugation patterns control the

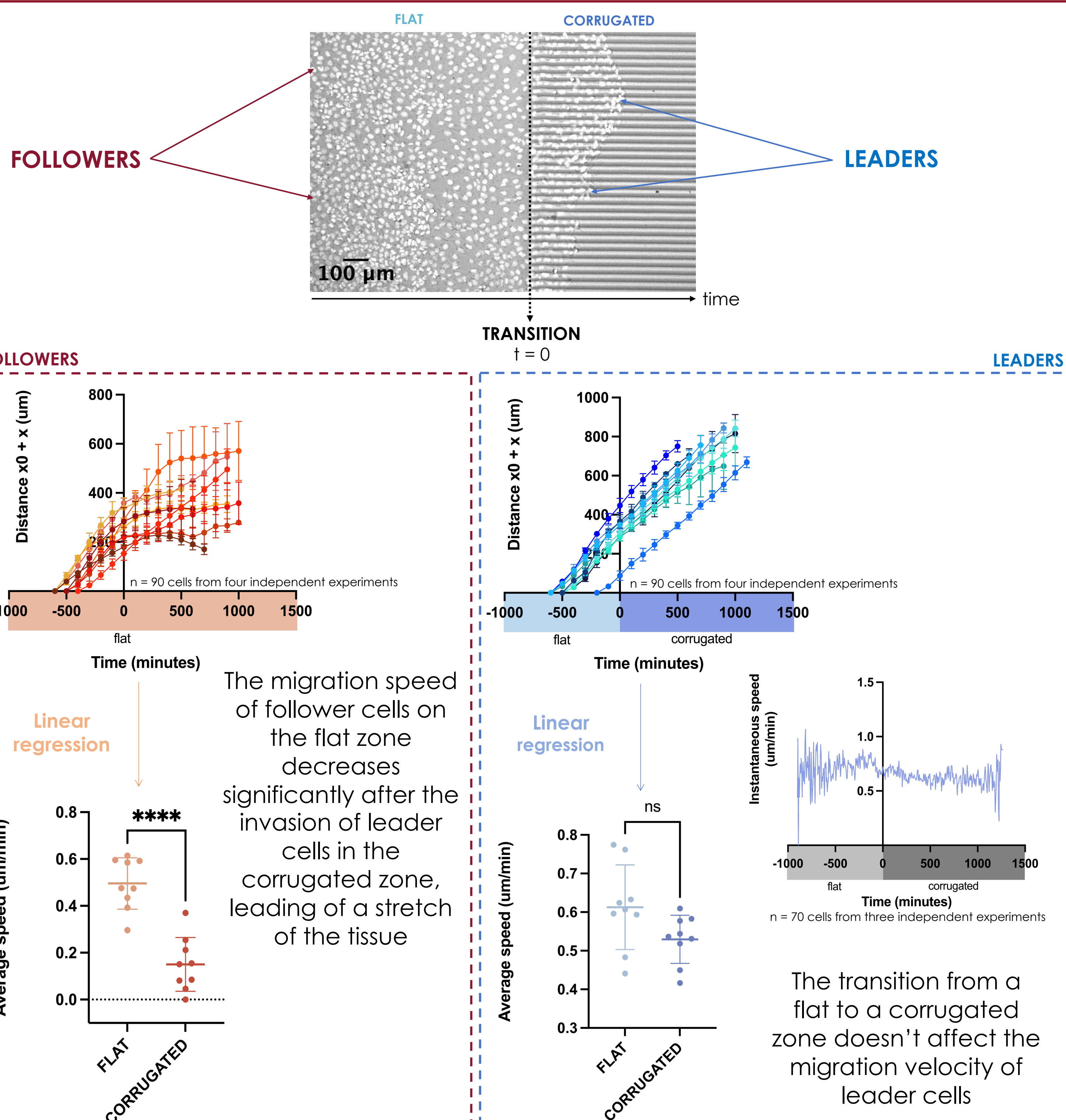
# UV photo-polymerization of hydroxy-PAAm hydrogels through an optical photomask



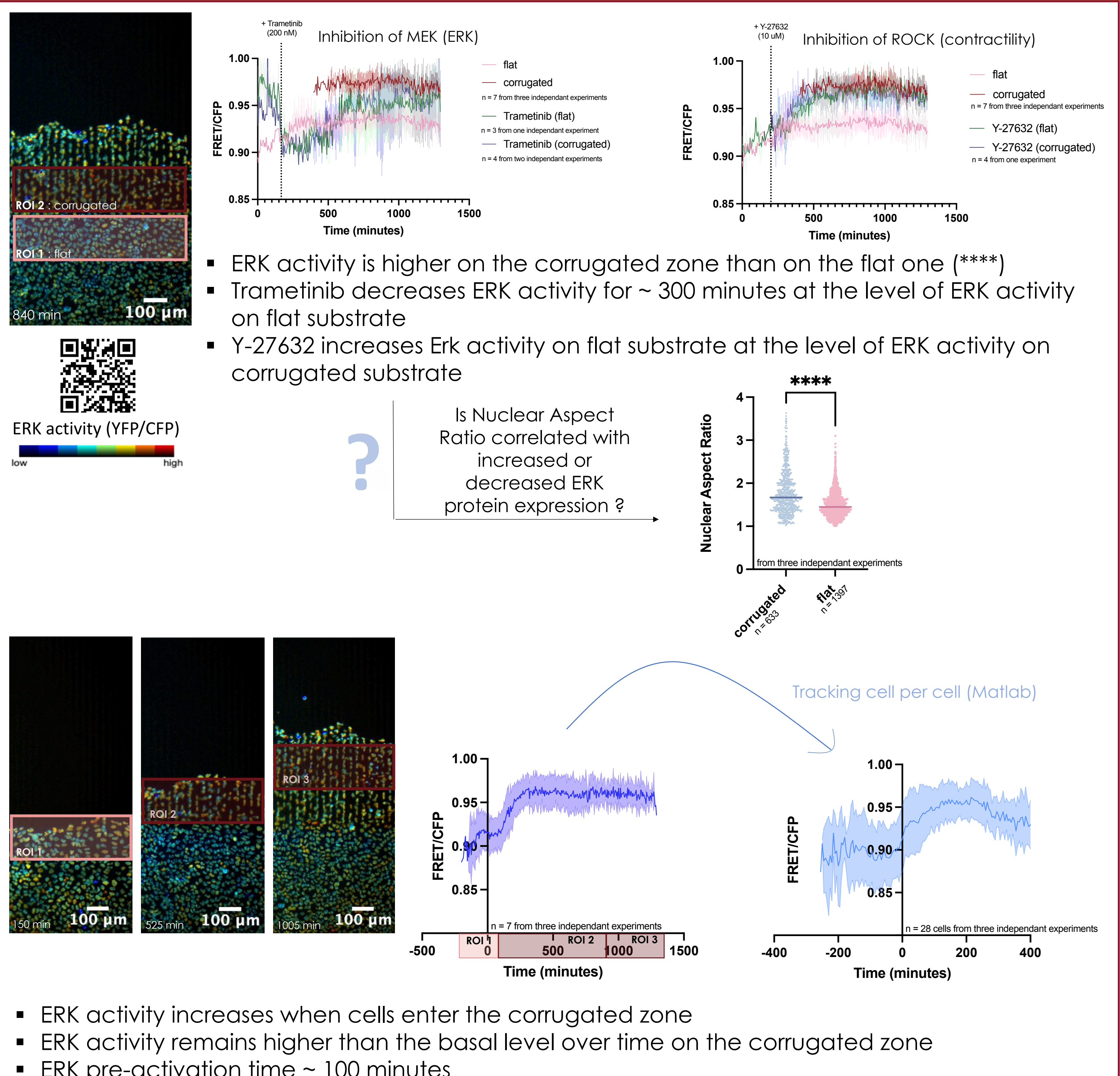
The epithelial flow is oriented parallel to the corrugations, while the tissue on flat zone shows isotropic flows and vortices



*Followers slow down after leaders invade corrugations*



*Erk activity: transition from a flat to a corrugated zone*



**Acknowledgement : Zoé Barbier is financially supported by FRIA-FNRS.**